Angewandte Chemie
Eine Zeitschrift der Gesellschaft Deutscher Chemiker

Supporting Information

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Corrigendum for 10.1002/anie.200461267 and 10.1002/ange.200461267

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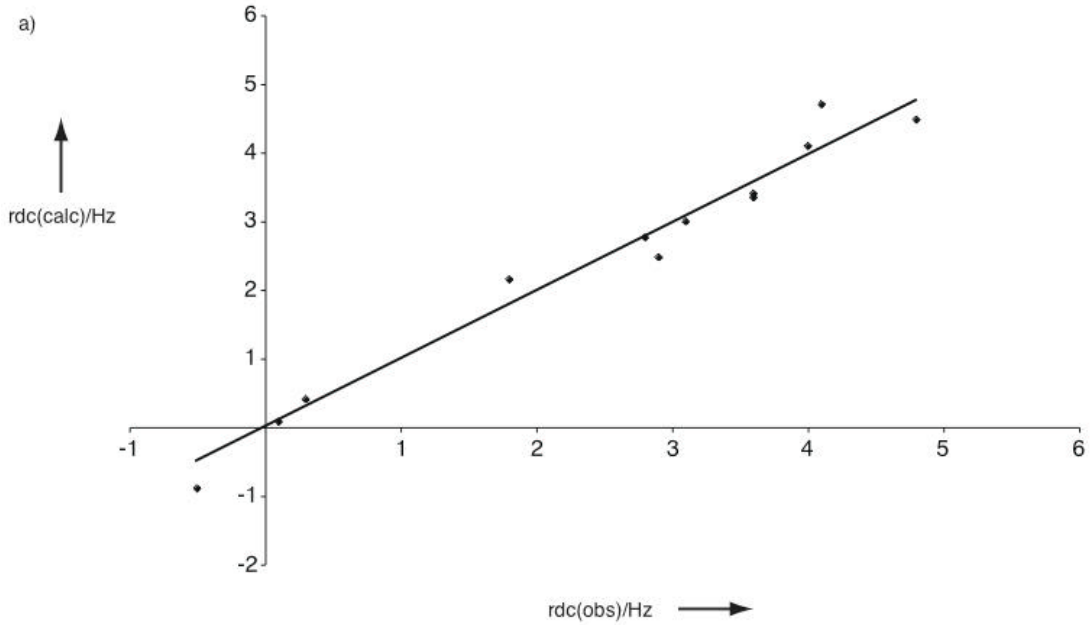

Figure SI-1: Correlation of calculated (calc) and experimental (obs) dipolar couplings for a 1 M (+)-menthol sample in a) DMSO with a correlation factor R = 0.98 and b) DMF (PH-gel concentration 12% (w/v)) with a correlation factor R = 0.96.

	1JCH / Hz	1DCH / Hz
C1-H1	123,6	4,1
C2-H2eq	123,7	2,8
C2-H2ax	127,4	3,6
C3-H3	136,4	4,8
C4-H4	125,0	4,0
C5-H5eq	117,8	3,1
C5-H5ax	126,4	3,6

C6-H6ax	126,4	3,6
C7-H7	124,3	1,8 (-0,5)
C8-H8	126,6	1,8
C9-H9	123,1	-0,5 (0,1)
C10-H10	124,0	-0,8 (0,3)

Table SI-1: Experimental ^{1}H-^{13}C scalar and dipolar couplings of (+)-menthol in DMSO recorded on a 400 MHz spectrometer at 298K. The value in brackets is according to Figure SI-1 the backcalculated $^{1}D_{CC}$ dipolar couplings.

All solutions used were degassed for at least 15-30 min to get rid of solubilized oxygen, that acts as an inhibitor during polymerization. This step is very important to ensure the homogeneity as well as the quality of the produced gels.

For 10 mL of pre-gel solution the amounts of the reactants are listed in Table S.1. AMPS, DMAA and BIS are dissolved in 9 ml of water. In some cases, it was necessary to slightly heat the solution to dissolve all the BIS. After the solution cooled down to room temperature 1 mL of APS stock solution was added and carefully mixed. The pre-gel solution is inserted into a gel cylinder

2-acrylamido-2-methyl-1-propanesulfonic acid (AMPS)	757 mg
N,N-dimethylacrylamide (DMAA)	377 µl
N,N'-methylenbisacrylamide (BIS)	30 mg
APS stock solution (0.015 g/l)	1 mL

Table SI-3 Composition of the PH-gel pre-gel solution.

with an inner diameter of 5.4 mm and polymerized for 2 h at 70°C in a heatable dessicator. Extreme care has to be taken when extracting the gel out of the cylinder with the screwable plunger, because it is very fragile at this stage of the procedure. It can take up to 10 min to extract one gel and it is advantageous to keep its top end lubricated with water. The PH-gel is washed once in an 0.02 M sodium hydroxide solution to neutralize its charges. This washing step turned out to be optional as gels not being washed in sodium hydroxide solution were swelling equally well. Then the gel is washed three times with water, each time for at least two hours and once overnight, to remove unpolymerized monomers and the starter APS. The swollen gel is then cut with a scalpel into a 3.7 cm long piece, which corresponds to a filling height of 500 µl in a 5 mm NMR-tube. Only the most homogeneous part of the gel is selected, avoiding parts that bear bubbles or other deficiencies. The cut gel is pierced in the middle through its long axis with a glass

capillary (diameter 0.3-0.4 mm), which is later used to position the gel in the middle of the fill height in the NMR-tube. The drying is carried out at room temperature on a petri dish covered with regular household foil to reduce friction between the gel and the glass surface. The gel is usually completely dry after 24 h. Drying at higher temperatures and reduced pressure is also possible, but bears the risk of cracking the gel. For NMR measurements the dried gel is put in a 5-mm NMR tube, that has been treated with Repelsilan (Sigma-Aldrich), which renders the glass surface hydrophobic and therefore facilitates the equilibration of the gel. With the help of the capillary the gel can be placed in the middle of the fill height. Then, one of the compatible solvents DMSO, DMF or water containing the compound of interest is added and the NMR tube is then sealed with a Shigemi plunger. If no Shigemi plunger is used, the gel tends to rip on top. After five days the gel has reswollen and is ready for use. If possible, equilibration times of 7-10 days are recommended.