



## Supporting Information

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## Quadruple and double helices of 8-fluoroquinoline oligoamides

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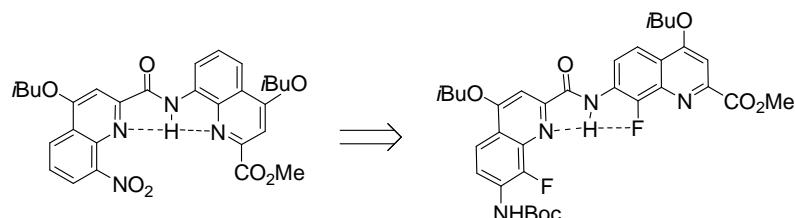
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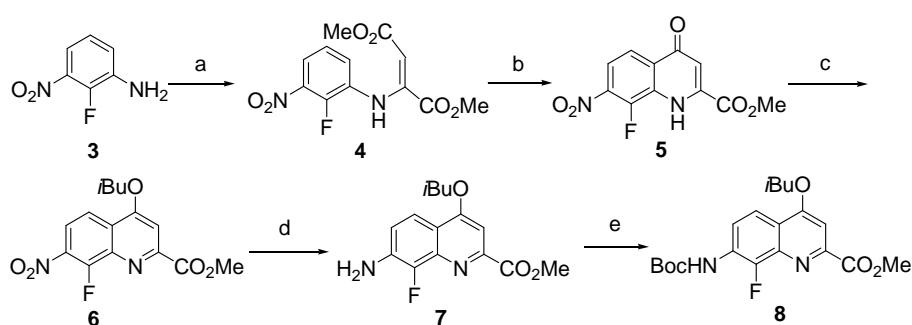
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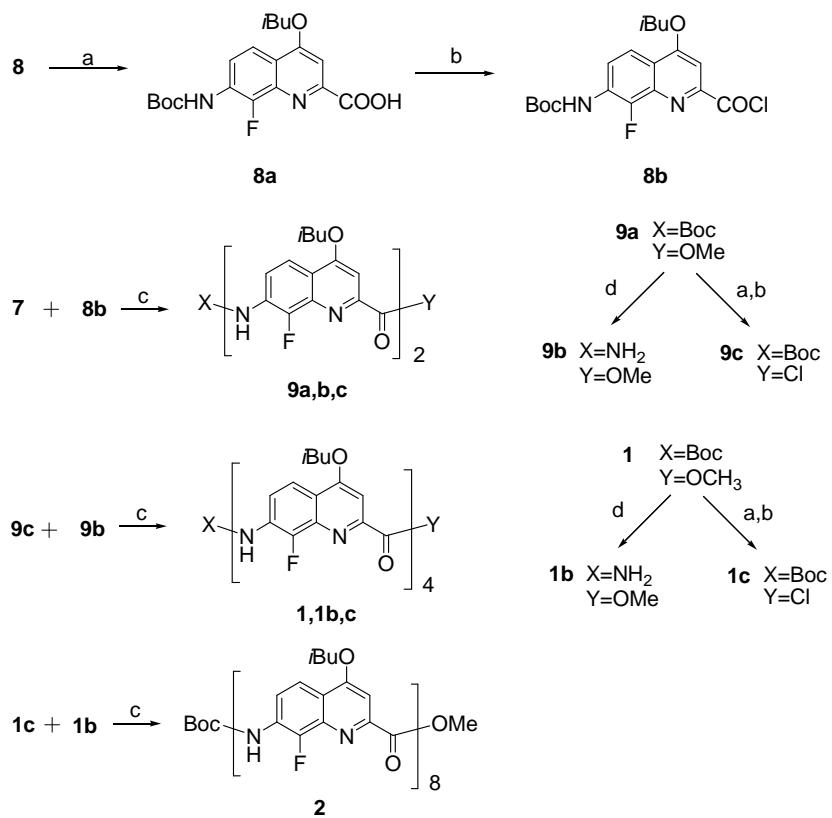
### Synthetic schemes



**Scheme S1** Expected change in curvature upon shifting the amine function from position 8 to position 7 in 2-quinolinicarboxamide oligomers.

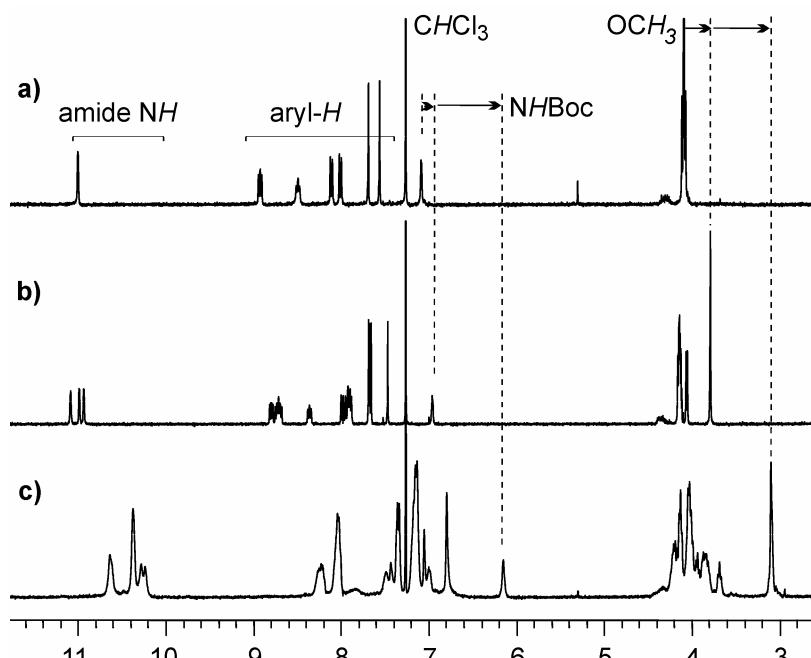


**Scheme S2.** Synthesis of fluoroquinoline monomer **8**: a) dimethylacetylene dicarboxylate, MeOH, Δ; b) diphenyl ether, reflux; c) *i*BuOH, DIAD, PPh<sub>3</sub>, THF, r.t.; d) Fe, AcOH, 120°C; e) di-*tert*-butyl dicarbonate, dioxane, heating for 7days.

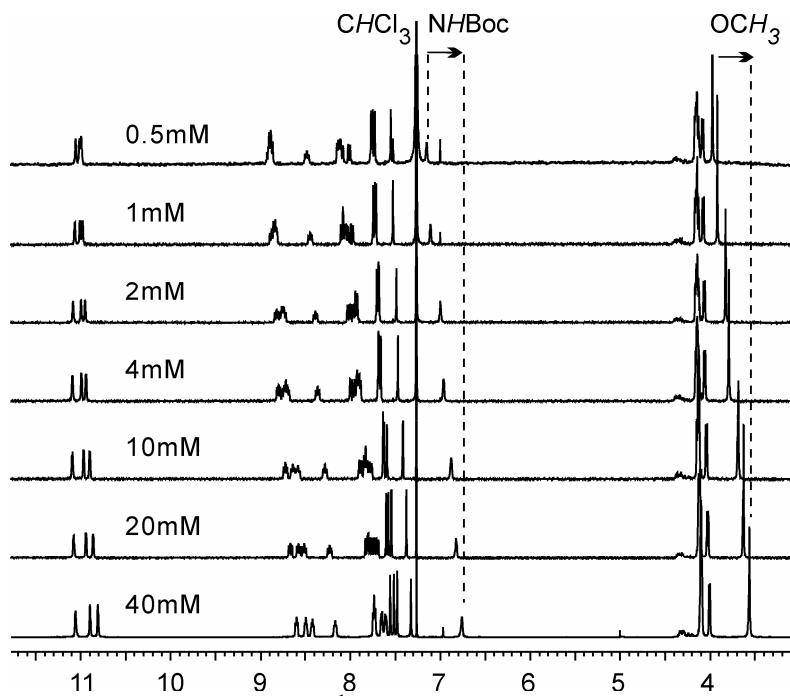


**Scheme S3.** Synthesis of oligofluoroquinoline amides. a) NaOH, THF/water, rt; b) 1-chloro-N,N,2-trimethylpropenylamine, DCM, rt, 2h; c) DIEA, DCM; d) TFA/DCM, rt.

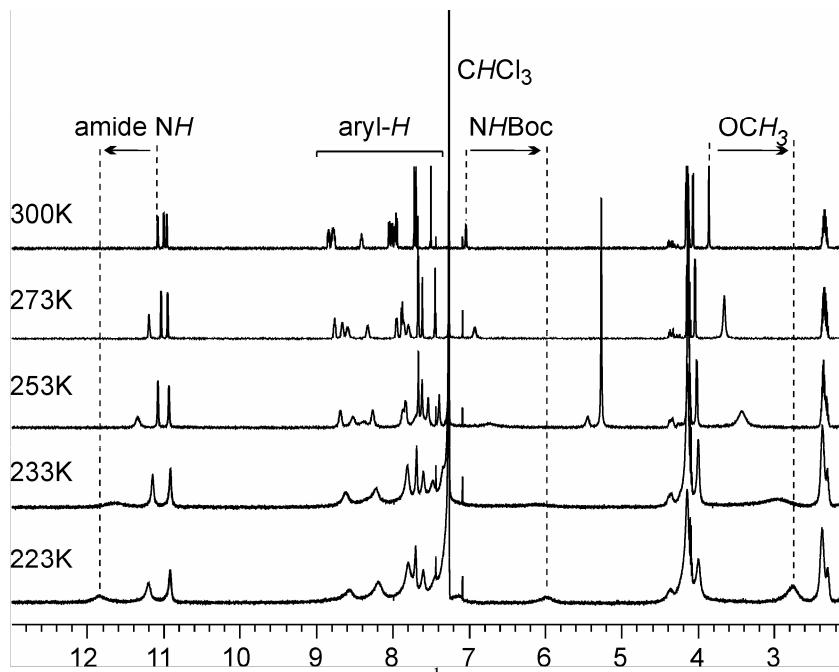
### NMR studies of aggregation



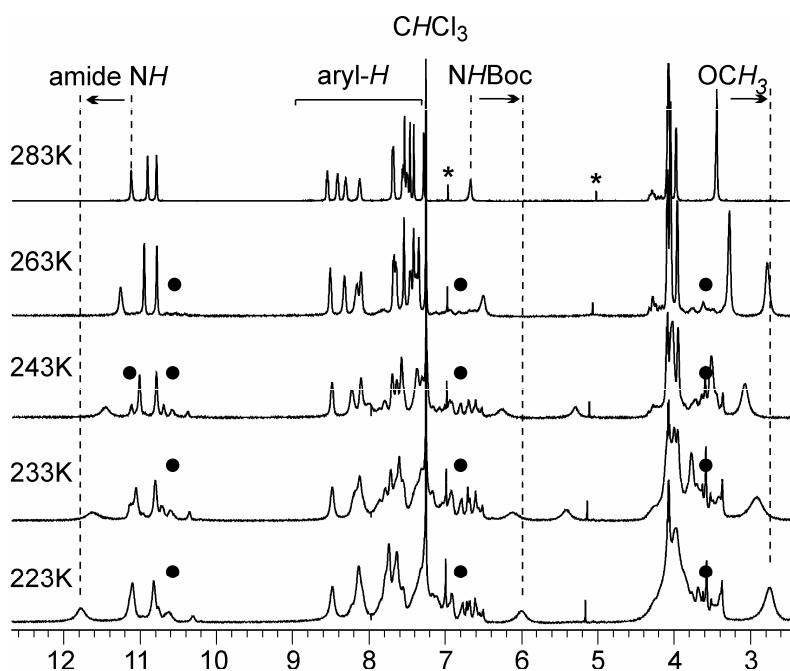
**Figure S1:** 400 MHz  $^1\text{H}$  NMR spectra of a) dimer **9a**, b) tetramer **1**, c) octamer **2** in  $\text{CDCl}_3$  (2mM) at 298K.



**Figure S2** Part of 400MHz  $^1\text{H}$  NMR of tetramer **1** at various concentrations in  $\text{CDCl}_3$  at 25°C.



**Figure S3** Temperature dependent  $^1\text{H}$  NMR spectra (600 MHz) of tetramer **1** (2mM) in  $\text{CDCl}_3$ . The sharp signal at 5.4 ppm observed at 253K is not identified but likely associated with the solvent or water. It appears in the 243-253K range in the spectra of **1** and in the spectra of **2**.

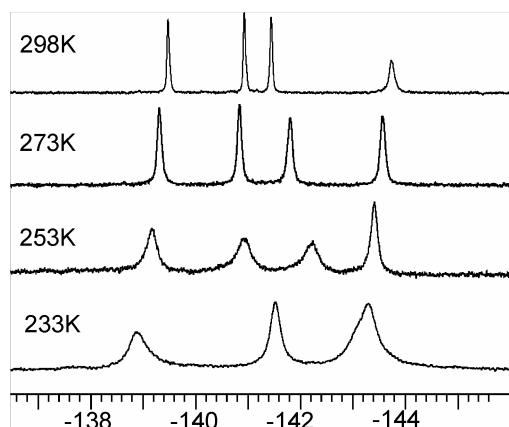


**Figure S4** Temperature dependent  $^1\text{H}$  NMR spectra (600 MHz) of tetramer **1** (40 mM) in  $\text{CDCl}_3$ . The black circles indicate signals of a new species appearing at low temperature. The stars indicate solvent signals

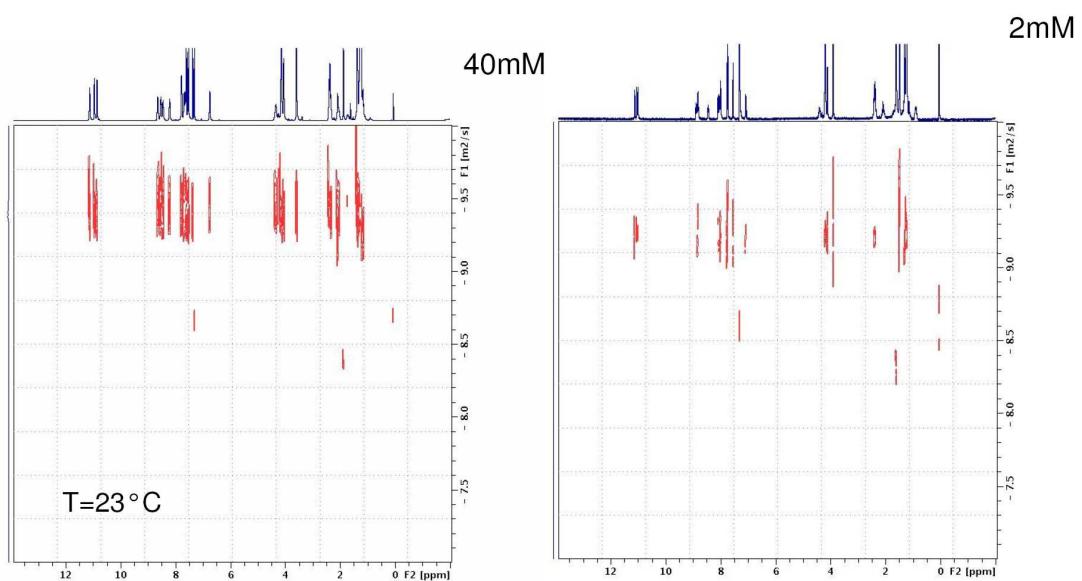
**Table S1** Changes of chemical shifts of tetramer **1** with various concentrations at 298K and 223K.

	$\Delta\delta_{\max}^{\text{a}}$ (BocNH)	$\Delta\delta_{\max}$ ( $\text{OCH}_3$ )	$\Delta\delta_{\max}$ (aromatic)
2 mM	-1.07	-1.09	-0.60
10 mM	-0.77	-0.80	-0.38
20 mM	-0.77	-0.85	-0.42
40 mM	-0.75	-0.80	-0.35

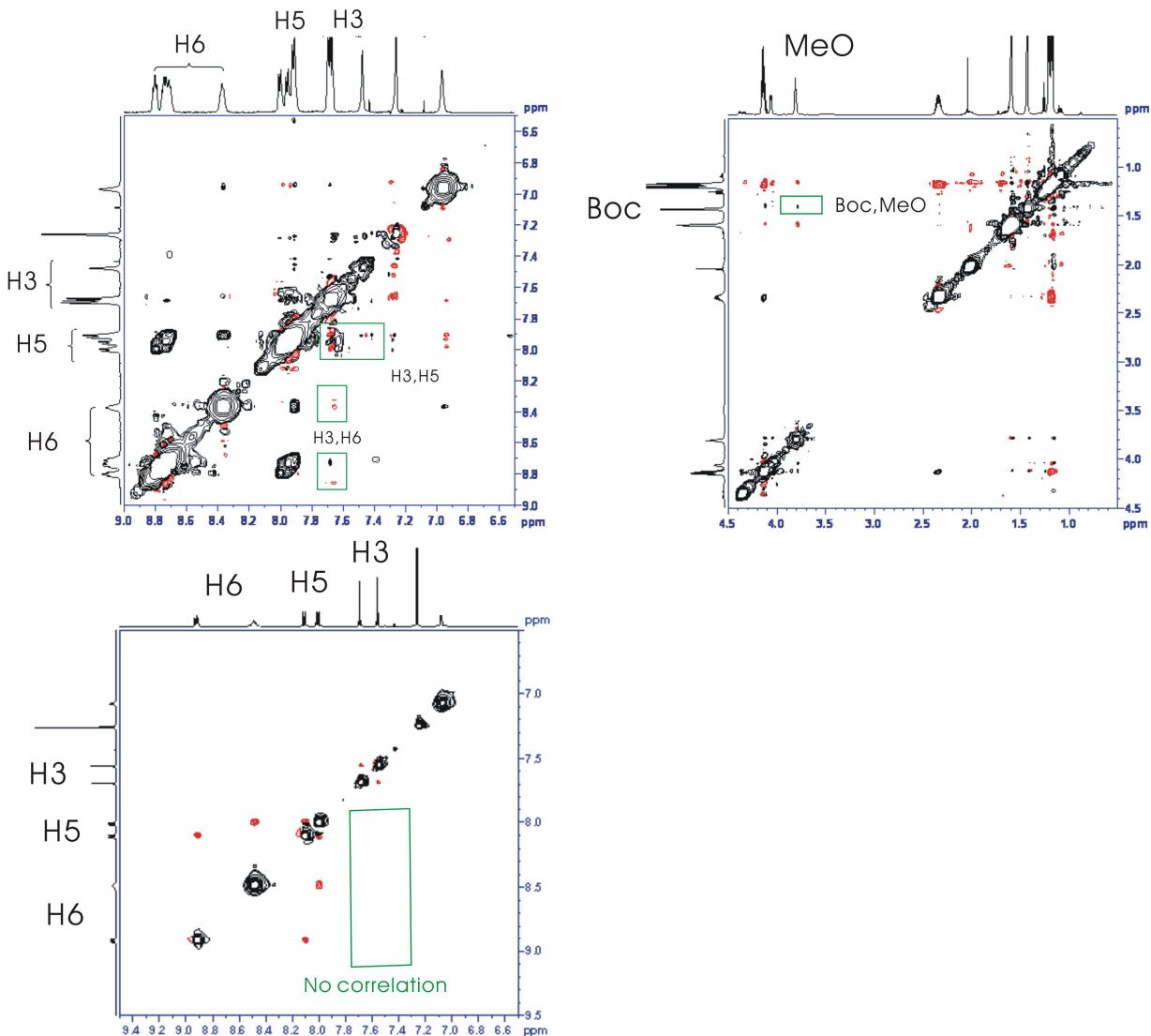
<sup>a</sup> $\Delta\delta_{\max} = \delta_{223\text{K}} - \delta_{298\text{K}}$



**Figure S5.** Temperature dependent  $^{19}\text{F}$  NMR spectra (400 MHz) of tetramer **1** (10mM) in  $\text{CDCl}_3$ .

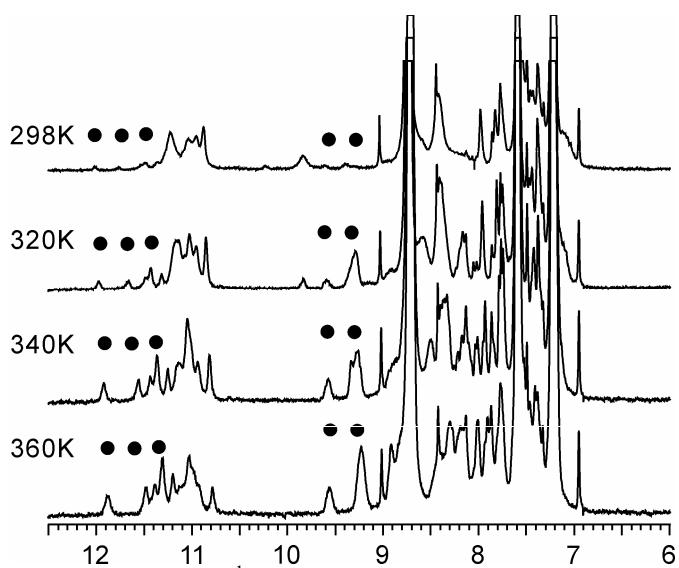


**Figure S6.**  $^1\text{H}$  DOSY NMR spectra (400 MHz) of tetramer **1** at 40 mM and 2 mM in  $\text{CDCl}_3$ .

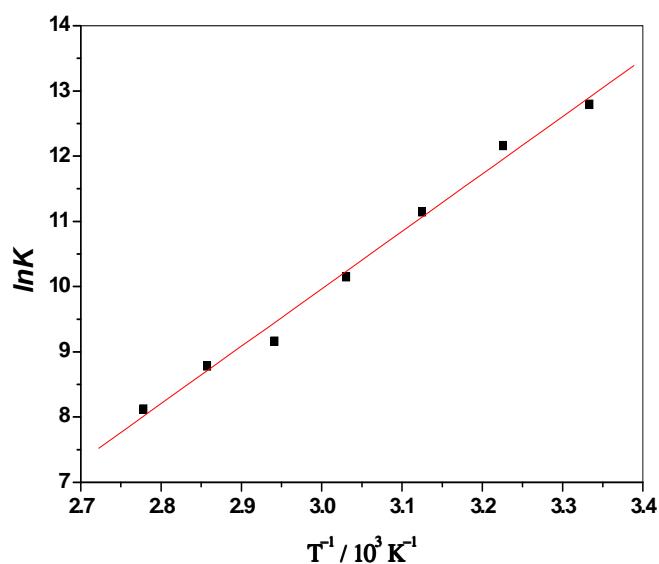


**Figure S7.** Partial NOESY spectra of tetramer **1** (top) and dimer **9a** (bottom) (4mM,  $\text{CDCl}_3$ , 600MHz). Correlations between H3 and H5/H6 are observed with **1** and not with **9a** suggesting that they correspond to intermolecular contacts associated with the aggregation of **1**, and not to

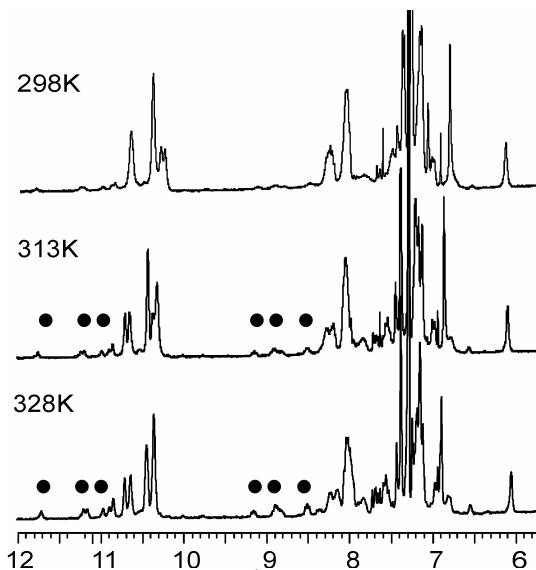
intramolecular correlations. The  $\text{NHBoc} - \text{CH}_3$  ester correlation in **1** (top right) is consistent both with the single helical conformation and with a head-to-tail arrangement of a duplex.



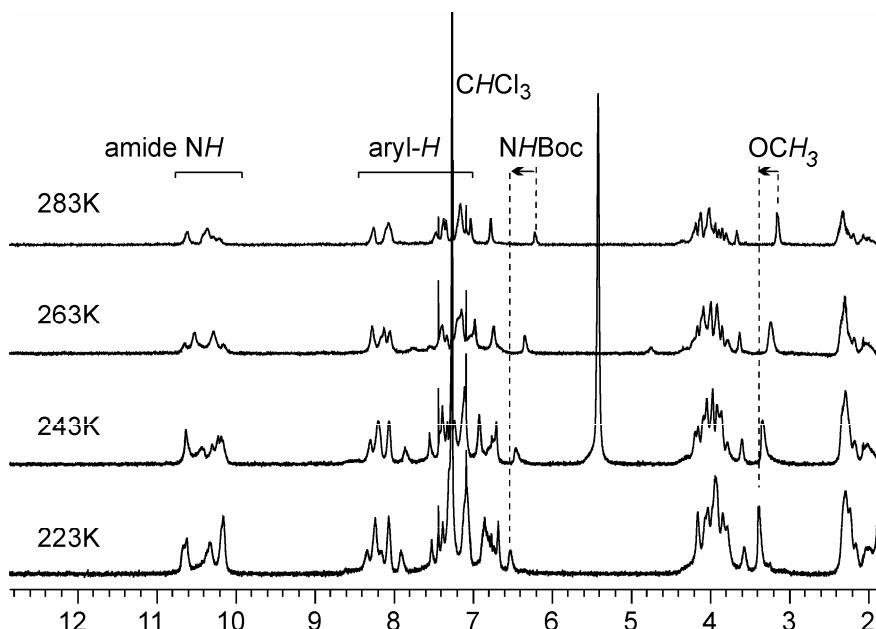
**Figure S8.** Partial  $^1\text{H}$  NMR spectra (300 MHz) of Octamer **2** (1mM) in pyridine-d<sub>5</sub> at various temperatures, showing a second set of signals appearing downfield (•).



**Figure S9.** Van't Hoff plot of the dimerization of octamer **2** in pyridine-d<sub>5</sub>. Experimental data were fitted to the Van't Hoff equation using linear regression analysis (red line,  $R=0.996$ ).  $\Delta H$  and  $\Delta S$  were extracted to be  $-73.2 \text{ kJ.mol}^{-1}$  and  $-136.3 \text{ J.K}^{-1}$ , respectively.



**Figure S10.** Partial  $^1\text{H}$  NMR spectra (400 MHz) of Octamer **2** (1mM) in  $\text{CDCl}_3$  at various temperatures, showing a second set of signals appearing downfield (•).



**Figure S11** Temperature dependence of  $^1\text{H}$  NMR spectra (400 MHz) of octamer **2** (1mM) in  $\text{CDCl}_3$ . The sharp signal at 5.4 ppm observed at 243K is not identified but likely associated with the solvent or water. It appears in the 243-253K range in the spectra of **1** and in the spectra of **2**.

## Experimental section

**General.** All reactions were carried out under a dry nitrogen atmosphere. NMR spectra were recorded on Bruker AVANCE 600 (600 MHz), Bruker DMX 300 (300 MHz) and Bruker AVANCE 400 (400 MHz) spectrometers. Chemical shifts are expressed in parts per million (ppm,  $\delta$ ) using residual solvent protons as internal standards (chloroform:  $\delta$  7.26 ppm; DMSO:  $\delta$  2.50 ppm). EI, ESI mass spectra and MALDI-TOF were obtained on GCT $\square$ LC-MS 2010 and Autotlex  $\square$  spectrometers, respectively.

**2-Fluoro-3-nitroaniline 3** To glacial acetic acid (80 mL) was added 2-Fluoro-1,3-dinitrobenzene<sup>[1]</sup> (4.98 g, 26.8 mmol). The mixture was heated to 120°C. Reduced iron powder (4.35 g, 2.9 equiv) was added in portions over a period of 40 min and the mixture was heated to reflux for 2.5 h. The hot reaction mixture was poured into cold water and extracted with ether (3×50 mL). The combined organic fractions were washed with brine (5×40 mL), dried over  $\text{Na}_2\text{SO}_4$ , filtered and evaporated to dryness to give 2.4 g (57.6% yield) of the crude product as a brown-black solid. Recrystallization from carbon tetrachloride afforded 1.95 g (46.8%) of pure 2-fluoro-3-nitroaniline, which was used without further purification.  $^1\text{H}$  NMR ( $\text{CDCl}_3$ ):  $\delta$

7.37-7.33 (m, 1H), 7.05-6.99 (m, 2H), 4.03 (s, 2H).<sup>[2]</sup> <sup>13</sup>C NMR (CDCl<sub>3</sub>): δ 146.0, 142.6, 137.2, 137.0., 124.1, 124.1, 121.5, 121.5, 124.2, 124.2. MS (EI): 156.

**Dimethyl N-(2-fluoro-3-nitro-phenyl)-amino-fumarate 4** To a solution of **3** (1.56 g, 0.010 mol) in MeOH (30 mL) was added dimethyl acetylenedicarboxylate (1.36 mL, 0.011 mol). After 18 h of stirring, the mixture was heated to reflux for 6 h and cooled. The resulting yellow prisms were collected by filtration to yield 2.0 g (67%) of the product. <sup>1</sup>H NMR (CDCl<sub>3</sub>): δ 9.64 (s, 1H), 7.74-7.70 (m, 1H), 7.19-7.12 (m, 2H), 5.72 (s, 1H), 3.78 (s, 6H). <sup>13</sup>C NMR (CDCl<sub>3</sub>): δ 169.5, 163.4, 149.3, 146.7, 145.5, 138.1, 138.1, 131.3, 131.2, 127.2, 127.2, 123.8, 123.7, 120.3, 120.3, 97.7, 53.1, 51.7. MS (ESI) : 299.2 [M+H]<sup>+</sup>, 321.1 [M+Na]<sup>+</sup>, 337.1 [M+K]<sup>+</sup>.

**Methyl 8-fluoro-7-nitro-4-(1*H*)-quinolone-2-carboxylate 5** A mixture of **4** (3.00 g, 0.010 mmol) and diphenyl ether (30.00 g) was heated with stirring at 240-250°C for 10 min. The product began to precipitate in the hot reaction medium. After cooling, the mixture was diluted with 100 ml of light petroleum ether to complete the precipitation. The solid was filtered off, washed well with hexane, and recrystallized from methanol in which it is sparingly soluble yield to give 2.3 g (86%) of the product. <sup>1</sup>H NMR (DMSO-d6): δ 12.85-12.56 (br, 1H), 8.20-8.10 (m, 3H), 3.96 (s, 3H). MS (ESI) : 267 [M+H]<sup>+</sup>, 289 [M+Na]<sup>+</sup>.

**Methyl 8-fluoro-4-isobutoxy-7-nitro-2-quinolinecarboxylate 6** A mixture of **5** (3.64 g, 13.7 mmol) and triphenylphosphine (3.95 g, 1.1 equiv.), and 2-methyl-1-propanol (1.52 mL, 1.2 equiv.) in anhydrous THF (40 mL) was cooled to 0°C under nitrogen. Diisopropyl azodicarboxylate (2.96 mL, 1.1 equiv.) was added dropwise and the mixture was stirred at 0°C for 30 min, then at room temperature for 2 h. The solvent was evaporated. The product was purified by flash chromatography (SiO<sub>2</sub>) eluting with CH<sub>2</sub>Cl<sub>2</sub> to yield 3.61 g (82%) of a white solid. <sup>1</sup>H NMR (CDCl<sub>3</sub>): δ 8.10-8.04 (m, 1H), 7.68 (s, 1H), 4.08 (d, J=16.4 Hz, 2H), 4.06 (s, 3H), 2.53-2.27 (m, 1H), 1.14 (d, J=6.8 Hz, 6H). <sup>13</sup>C NMR (CDCl<sub>3</sub>): δ 165.4, 162.2, 162.8, 153.9, 151.6, 151.1, 139.5, 139.4, 136.6, 126.5, 121.8, 121.8, 118.0, 117.9, 104.0, 76.1, 53.6, 28.2, 19.2. MS (ESI) : 323.2 [M+H]<sup>+</sup>.

**Methyl 7-amino-8-fluoro-4-isobutoxy-2-quinolinecarboxylate 7.** The nitro quinoline **6** (0.32 g, 1.0 mmol) was dissolved in glacial acetic acid (9 mL) and the solution was heated to 120°C Reduced iron powder (0.17 g, 3.0 mmol) was added in one portion and the mixture was stirred at 120°C for 2 h. The hot mixture was poured into water, and the precipitate was filtered off and washed with cold water to give 0.23 g (80%) of the product as a yellow solid which was used without additional purification. <sup>1</sup>H NMR (CDCl<sub>3</sub>): δ 7.86 (d, J=11.9 Hz, 1H), 7.41 (s, 1H), 7.10 (t, J=11.1 Hz, 1H), 4.15 (br, 2H), 4.04 (s, 3H), 4.02 (d, J=6.6 Hz, 2H), 2.30-2.24 (m, 1H) 2.53-2.27 (m, 1H), 1.12 (d, J=6.6 Hz, 6H). <sup>13</sup>C NMR (CDCl<sub>3</sub>): δ 166.5, 162.8, 149.7, 145.6, 142.3, 139.8, 139.7, 135.0, 134.8, 119.7, 117.7, 117.7, 116.0, 99.1, 75.1, 53.3, 28.2, 19.3. MS (ESI): 293.1 [M+H]<sup>+</sup>, 315.2 [M+Na]<sup>+</sup>, 331.1 [M+K]<sup>+</sup>.

**Methyl 7-(tert-butoxycarbonylamino)-8-fluoro-4-isobutoxy-2-quinolinecarboxylate 8** A solution of **7** (0.44 g, 1.5 mmol) in dioxane (30 mL) containing di-*tert*-butyl-dicarbonate (1.64 g, 7.5 mmol) was heated at 95°C for 7 days. The solvent was removed in vacuo, and the residue was purified by flash chromatography on silica gel eluting with CH<sub>2</sub>Cl<sub>2</sub> to afford the pure product (0.56 g, 95%). <sup>1</sup>H NMR (CDCl<sub>3</sub>): δ 8.50 (t, J=8.0 Hz, 1H), 7.98 (d, J=9.3 Hz, 1H), 7.51 (s, 1H), 7.05 (d, J=1.2 Hz, 1H), 4.06 (s, 3H), 4.05 (d, J=6.9 Hz, 2H), 2.32-2.26 (m, 1H), 1.57 (s, 9H), 1.14 (d, J=6.6 Hz, 6H). <sup>13</sup>C NMR (CDCl<sub>3</sub>): δ 166.0, 162.6, 152.2, 149.8, 146.9, 144.4, 138.6, 138.5, 127.6, 127.5, 120.2, 118.6, 117.4, 100.3, 81.5, 75.2, 53.2, 28.2, 28.1, 19.2. MS (ESI): 393.2 [M+H]<sup>+</sup>.

**Dimer 9a.** Monomer **8** (1.35 g, 3.44 mmol) was dissolved in a mixture of THF (60 mL) and H<sub>2</sub>O (6 mL). To this solution was added NaOH (0.344 g, 2.5eq). The solution was stirred at room temperature for 5h. Then solution was neutralized with 1N HCl to pH = 4~5, and concentrated under reduced pressure to remove THF. H<sub>2</sub>O (30 mL) was added to the residue. The aqueous phase was extracted with CH<sub>2</sub>Cl<sub>2</sub> (3×50 mL). The combined organic phases were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, then evaporated to give monomer acid **8a** as a yellowish solid. It was dried in vacuo, and then dissolved in CH<sub>2</sub>Cl<sub>2</sub> (40 mL). 1-chloro-*N,N*,2-trimethyl-propenylamine (0.92 g, 2 .0 eq) was added. The reaction mixture was stirred at RT for 2 h resulting in a homogeneous solution, and then evaporated to provide the corresponding acid chloride **8c**. To a solution of the monomer amine **7** (0.95 g, 3.26 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (40 mL) containing DIEA (2.3 mL, 13.04 mmol) was added a solution of acide chloride **8c** in CH<sub>2</sub>Cl<sub>2</sub> (40 mL) via cannula. The reaction mixture was stirred at RT overnight. The solution was evaporated and the product was purified by flash chromatography (SiO<sub>2</sub>) eluting with CH<sub>2</sub>Cl<sub>2</sub>. Yield 2.01 g (95%) of a white solid. <sup>1</sup>H NMR (CDCl<sub>3</sub>): δ 11.00 (s, 1H), 8.94-8.90 (d, J=9.2 Hz, J=2.4 Hz 1H), 8.497 (m, 1H), 8.11 (d, J=9.3 Hz, 1H), 8.01 ((d, J=9.3 Hz, 1H), 7.68 (s, 1H), 7.56 (s, 1H), 7.09 (s, 1H), 4.10 (d, J=17.3 Hz, 4H), 4.08 (s, 3H), 2.34-2.28 (m, 2H), 1.59 (m, 9H), 1.15 (t, J=6.2 Hz, 6H).

<sup>13</sup>C NMR (CDCl<sub>3</sub>): δ 165.9, 162.9, 162.9, 162.3, 162.2, 162.0, 152.1, 150.6, 149.6, 147.7, 146.4, 145.2, 143.8, 138.3, 138.2, 137.2, 137.1, 127.4, 127.3, 126.7, 126.6, 120.2, 119.7, 119.2, 118.3, 117.1, 100.3, 97.4, 81.4, 75.2, 75.1, 53.1, 28.2, 28.1, 28.0, 19.1. MS (ESI): 653.3 [M+H]<sup>+</sup>.

**Tetramer 1** Dimer **9a** (0.84 g 1.29 mmol) was dissolved in CH<sub>2</sub>Cl<sub>2</sub> (10 mL), and excess TFA (5 mL) was added. The mixture was stirred at room temperature for 3h. The solvent was evaporated and the residue was dissolved in CH<sub>2</sub>Cl<sub>2</sub> (20 mL), washed with saturated NaHCO<sub>3</sub>, dried over Na<sub>2</sub>SO<sub>4</sub> and then evaporated to give dimer amine **9b** as a yellow solid. It was dried in vacuo, and used without further purification. Separately, dimer **9a** (0.89 g, 1.37 mmol) was dissolved in a mixture of THF (50 mL) and H<sub>2</sub>O (5 mL). NaOH (0.137 g, 3.43 mmol) was added, and the solution was stirred at room temperature for 5h. The solution was neutralized with 1N HCl to pH = 4~5, then concentrated under reduced pressure to remove THF. H<sub>2</sub>O (30 mL) was added to the residue. The aqueous phase was extracted with CH<sub>2</sub>Cl<sub>2</sub> (3×50 mL). The combined organic phases were dried over Na<sub>2</sub>SO<sub>4</sub> and then evaporated to give the corresponding dimer acid as a yellowish solid. It was dried in vacuo, and then dissolved in CH<sub>2</sub>Cl<sub>2</sub> (40 mL). To this solution was added 1-chloro-*N,N*, 2-trimethylpropenylamine (0.36 g, 2.74 mmol). The reaction mixture was stirred at RT for 2 h resulting in a homogeneous solution, then evaporated to provide the corresponding acid chloride **9c**. To a solution of the dimer amine **9b** in CH<sub>2</sub>Cl<sub>2</sub> (40 mL) containing DIEA (0.89 mL, 5.16 mmol) was added a solution of acid chloride **9c** in CH<sub>2</sub>Cl<sub>2</sub> (40 mL) via cannula. The reaction mixture was stirred at RT overnight. The solution was evaporated and the product was purified by flash chromatography (SiO<sub>2</sub>) eluting with CH<sub>2</sub>Cl<sub>2</sub>/EtOAc, 20:1. Yield 1.39 g (92%) of a white solid. <sup>1</sup>H NMR (4 mM, CDCl<sub>3</sub>): δ 11.08 (s, 1H), 10.98 (s, 1H), 10.93 (s, 1H), 8.31-8.67 (m, 3H), 8.36 (t, J=6.9 Hz, J=2.0 Hz, 1H), 8.00-7.88 (m, 4H), 7.68 (s, 1H), 7.67 (s, 1H), 7.65 (s, 1H), 7.47 (s, 1H), 6.96 (s, 1H), 4.16-4.05 (m, 8H), 3.79 (s, 3H), 2.38-2.29 (m, 4H), 1.42 (s, 9H), 1.22-1.17 (m, 24H). <sup>13</sup>C NMR (CDCl<sub>3</sub>): δ 166.1, 163.0, 162.8, 162.2, 162.1, 152.0, 150.9, 150.7, 149.4, 148.1, 147.7, 147.3, 146.2, 145.6, 145.3, 144.8, 143.7, 138.4, 138.3, 137.2, 137.1, 136.9, 127.3, 127.2, 126.7, 126.6, 119.5, 119.4, 119.2, 119.0, 118.9, 118.3, 116.2, 116.6, 100.2, 97.9, 97.6, 81.2, 75.4, 75.3, 75.2, 52.8, 28.4, 28.3, 28.1, 19.4. MS (MALDI-TOF) : 1195.6 [M+Na]<sup>+</sup>, 1211.5 [M+K]<sup>+</sup>.

**Octamer 2** was prepared from tetramer 1 via saponification on one hand and TFA treatment on the other hand to get the tetramer acid and tetramer amine, respectively. The synthetic procedures are the same as used for the synthesis of **1**. Yield > 80%. <sup>1</sup>H NMR (4 mM, CDCl<sub>3</sub>): δ 10.65 (s, 1H), 10.61 (s, 1H), 10.37 (s, 3H), 10.31 (s, 2H), 8.22 (br, 2H), 8.03 (br, 4H), 7.85 (br, 1H), 7.50-7.34 (m, 5H), 7.14 (br, 7H), 7.05 (s, 1H), 6.98 (br, 1H), 6.80 (s, 3H), 6.14 (s, 1H), 4.41-3.67 (m, 16H), 3.10 (s, 3H), 2.41-2.26 (m, 8H), 1.33-1.11 (m, 41H). <sup>13</sup>C NMR (CDCl<sub>3</sub>): δ 165.5, 162.1, 161.9, 161.7, 161.6, 161.4, 161.3, 161.1, 160.7, 151.6, 150.5, 150.2, 149.5, 149.3, 148.9, 148.1, 146.8, 146.6, 146.3, 145.3, 144.2, 144.0, 143.7, 142.8, 136.8, 136.4, 136.1, 136.0, 135.7, 126.2, 126.1, 125.8, 118.6, 118.4, 118.0, 117.8, 117.6, 117.5, 117.3, 115.6, 99.3, 98.0, 97.7, 97.5, 97.3, 96.8, 80.9, 75.1, 74.9, 74.7, 52.2, 28.5, 28.4, 28.3, 28.2, 27.8, 19.6, 19.6, 19.5, 19.5, 19.4, 19.3, 19.3, 19.2, 19.2, 19.1, 19.1. MS (MALDI-TOF) : m/z 2236.8 [M +H+ Na]<sup>+</sup>, 2252.7 [M+H+K]<sup>+</sup>

### X-ray crystallography

Data were collected using a Rigaku Rapid diffractometer equipped with an MM007 microfocus rotating anode generator with monochromatized Cu-K<sub>α</sub> radiation (1.54178 Å) and varimax optics. The data collection, unit cell refinement and data reduction were performed using the CrystalClear software package. The positions of non-H atoms were determined by the program SHELXS 87 and the position of the H atoms were deduced from coordinates of the non-H atoms and confirmed by Fourier Synthesis. H atoms were included for structure factor calculations but not refined.

Single crystals of **1** (single helix), [C<sub>62</sub>H<sub>64</sub>N<sub>8</sub>O<sub>11</sub>F<sub>4</sub>(C<sub>6</sub>H<sub>5</sub>Cl)(H<sub>2</sub>O)<sub>3</sub>], were grown from chlorobenzene/n-hexane. A single colorless crystal of dimensions 0.3 × 0.2 × 0.2 mm was selected, mounted on a cryoloop under oil and frozen into a N<sub>2</sub> stream at 223K. Crystals belong to the Triclinic space group P-1 with unit cell dimensions: a = 13.2847 (17) Å, b = 15.545 (2) Å, c = 16.933 (2) Å, α = 101.885 (3)°, β = 99.133 (5)°, γ = 97.076 (4)°, V = 3334.3 (7) Å<sup>3</sup>, and Z = 2 (FW is 1333.76, ρ = 1.328 Mg m<sup>-3</sup>). Reflections were collected from 2.72° ≤ θ ≤ 72.83° for a total of 45852 of which 12240 were unique (*R*<sub>int</sub> = 0.2385) having I > 2σ (I); number of parameters is 859. Final *R* factors were R<sub>1</sub> = 0.1477 (I > 2σ (I)), wR<sub>2</sub> = 0.4603 (all data), GOF = 1.356 from SHELX, maximal residual electron density is 0.638 e Å<sup>-3</sup>. CCDC deposition #665055

Single crystals of **1** (quadruple helix), [C<sub>135</sub>H<sub>133</sub>N<sub>16</sub>O<sub>35.55</sub>Cl<sub>2</sub>F<sub>8</sub>], were grown from toluene/1,2-dichloroethane/n-hexane. A single colorless crystal of dimensions 0.1 × 0.2 × 0.3 mm was selected, mounted on a cryoloop under oil and frozen into a N<sub>2</sub> stream at 223K. Crystals belong to the Orthorhombic space group Fdd2 with unit cell dimensions: a = 45.7987 (6) Å, b = 40.4084 (5) Å, c = 40.8397 (4) Å, α = 90°, β = 90°, γ = 90°, V = 75580 (2) Å<sup>3</sup>, and Z = 48 (FW is 2771.46, ρ = 0.974 Mg m<sup>-3</sup>). Reflections were collected from 2.90° ≤ θ ≤ 73.23° for a total of 177150 of which 35050 were unique (*R*<sub>int</sub> = 0.1385) having I > 2σ (I);

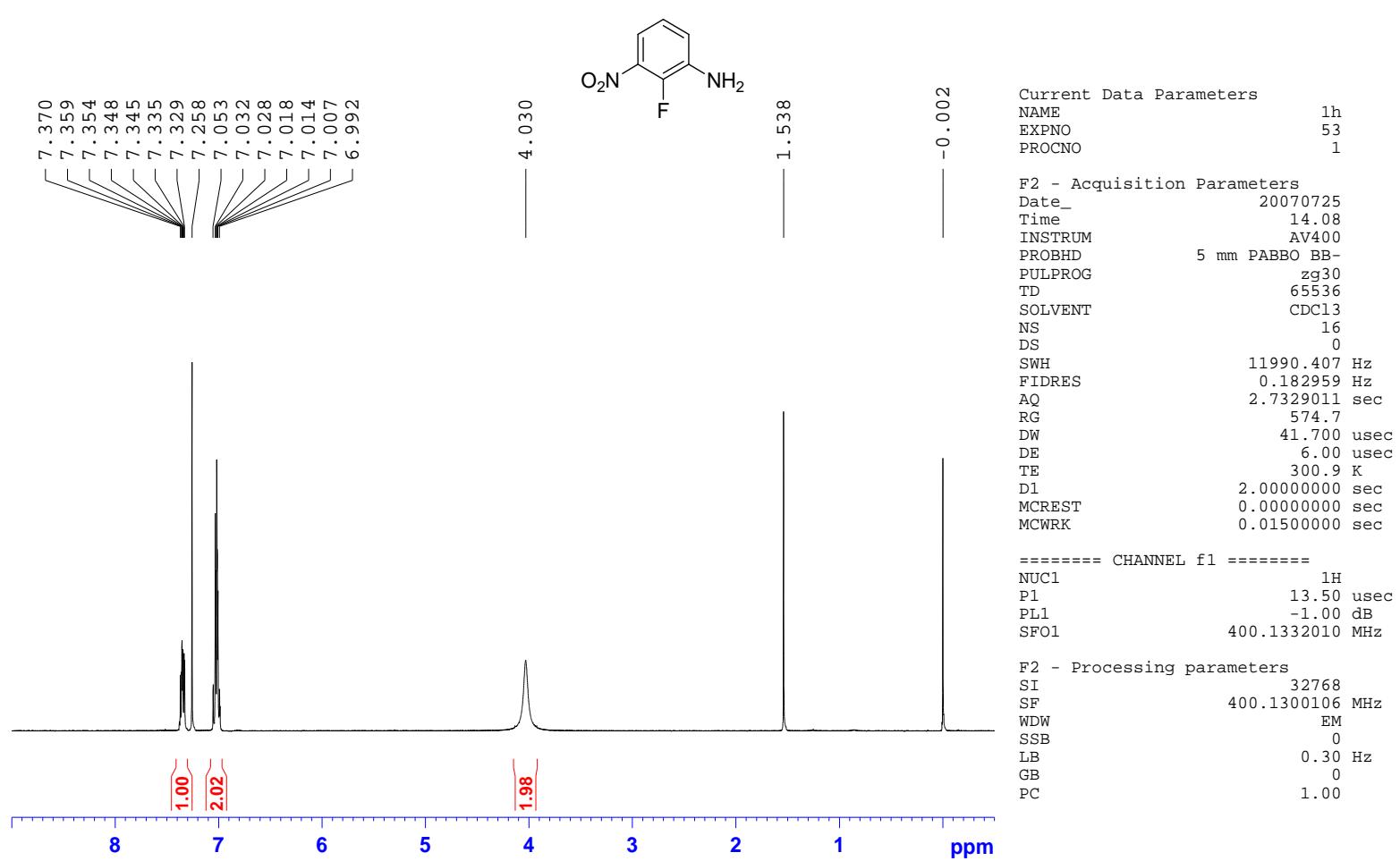
number of parameters is 1764. Final  $R$  factors were  $R_1 = 0.1653$  ( $I > 2\sigma(I)$ ),  $wR_2 = 0.4430$  (all data), GOF = 1.352 from SHELX, maximal residual electron density is 0.432 e Å<sup>-3</sup>. CCDC deposition #665054

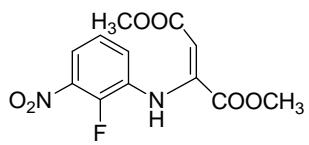
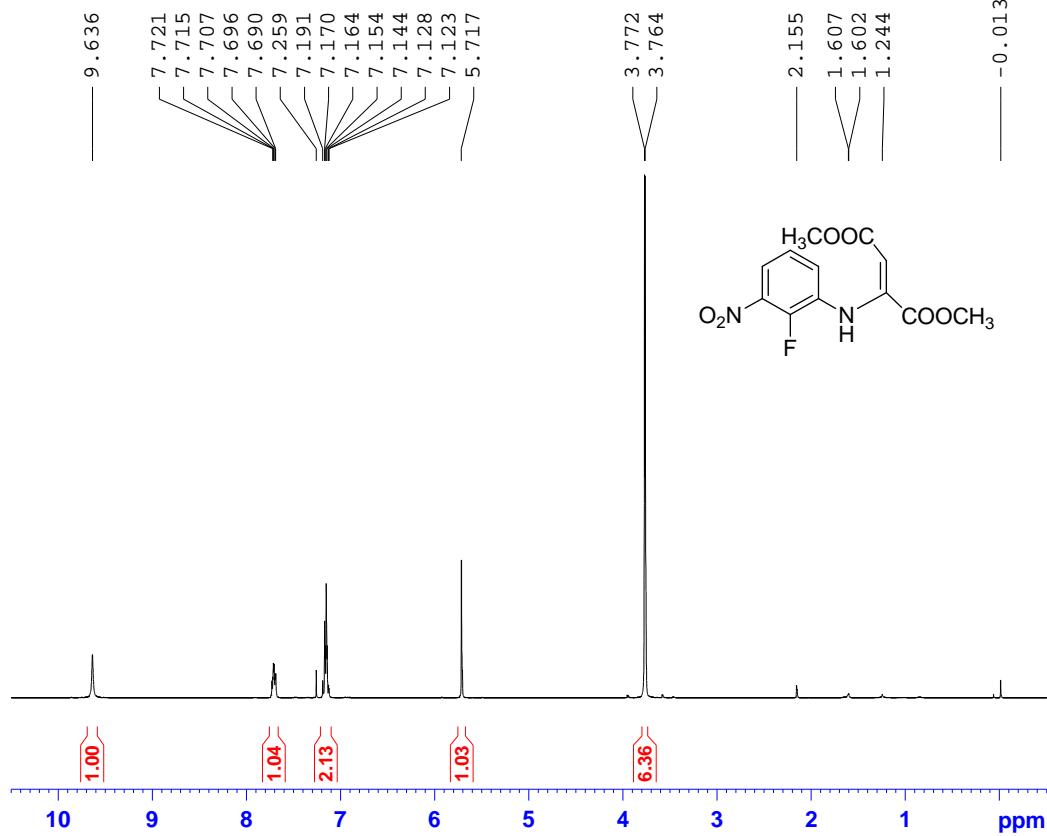
Single crystals of **2** (double helices), [C<sub>944</sub>H<sub>928</sub>N<sub>128</sub>O<sub>152</sub>F<sub>64</sub>], were grown from 1,2 dichloroethane/n-hexane. A single colorless crystal of dimensions 0.1 × 0.2 × 0.2 mm was selected, mounted on a cryoloop under oil and frozen into a N<sub>2</sub> stream at 223K. Crystals belong to the triclinic space group P-1 with unit cell dimensions:  $a = 25.391$  (13) Å,  $b = 26.509$  (13) Å,  $c = 27.909$  (17) Å,  $\alpha = 85.87$  (3)°,  $\beta = 63.32$  (3)°,  $\gamma = 62.56$  (3)°,  $V = 14680$  (2) Å<sup>3</sup>, and  $Z = 2$  (FW is 21511.84,  $\rho = 1.217$  Mg m<sup>-3</sup>). Reflections were collected from  $6.60^\circ \leq \theta \leq 72.30^\circ$  for a total of 153208 of which 47937 were unique ( $R_{\text{int}} = 0.1655$ ) having  $I > 2\sigma(I)$ ; number of parameters is 3503. Final  $R$  factors were  $R_1 = 0.1339$  ( $I > 2\sigma(I)$ ),  $wR_2 = 0.3812$  (all data), GOF = 0.660 from SHELX, maximal residual electron density is 0.695 e Å<sup>-3</sup>. CCDC deposition #665053

## References

- 1 J. F. K. Wilshire, *Aust. J. Chem.* **1967**, *20*, 2089–2811.
- 2 M. Hudlicky, H. M. Bell, *J. Fluorine Chem.* **1974**, *4*, 19–23.

<sup>1</sup>H and <sup>13</sup>C NMR spectra of all relevant synthetic intermediates and title compounds.



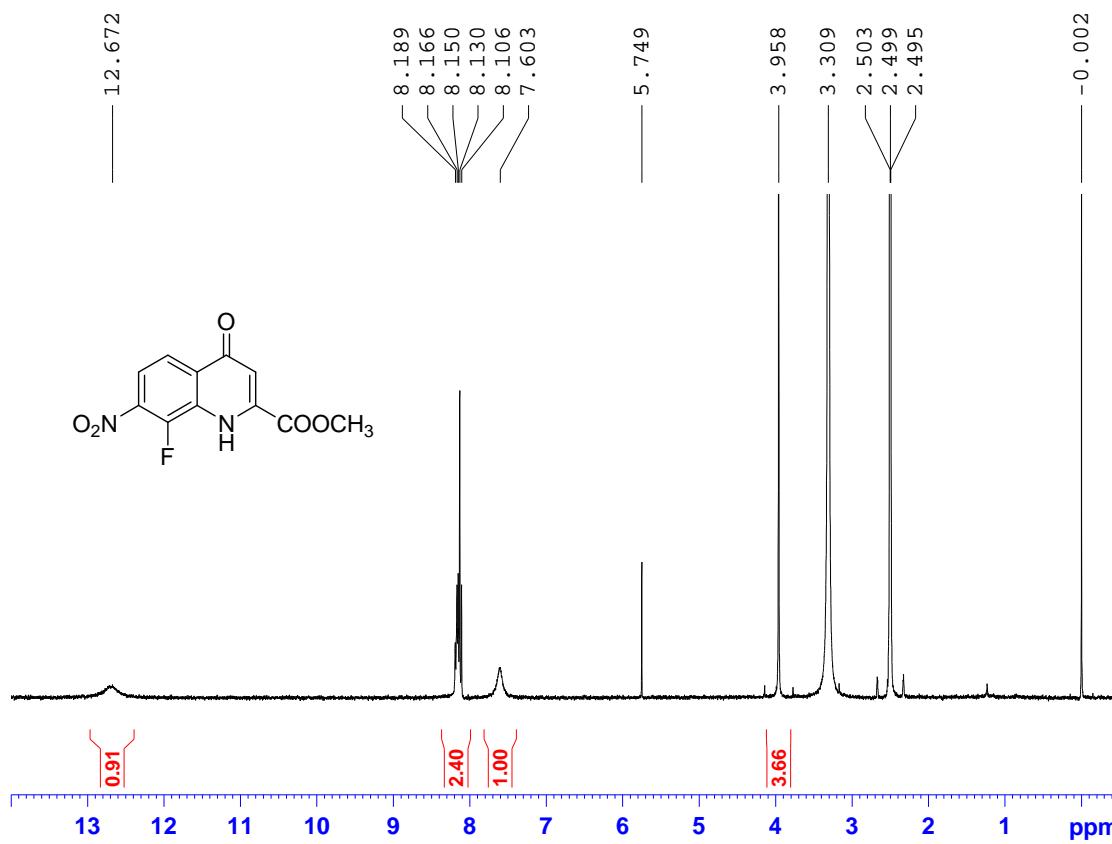


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 PROCNO 1

F2 - Acquisition Parameters  
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 PULPROG zg30  
 TD 65536  
 SOLVENT CDCl<sub>3</sub>  
 NS 8  
 DS 0  
 SWH 10822.511 Hz  
 FIDRES 0.165138 Hz  
 AQ 3.0278132 sec  
 RG 143.7  
 DW 46.200 usec  
 DE 6.00 usec  
 TE 300.6 K  
 D1 4.0000000 sec  
 MCREST 0.0000000 sec  
 MCWRK 0.0150000 sec

===== CHANNEL f1 =====  
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 P1 8.00 usec  
 PL1 -4.00 dB  
 SFO1 400.1332010 MHz

F2 - Processing parameters  
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 SF 400.1300092 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 0.10



Current Data Parameters

NAME	1h
EXPNO	76
PROCNO	1

F2 - Acquisition Parameters

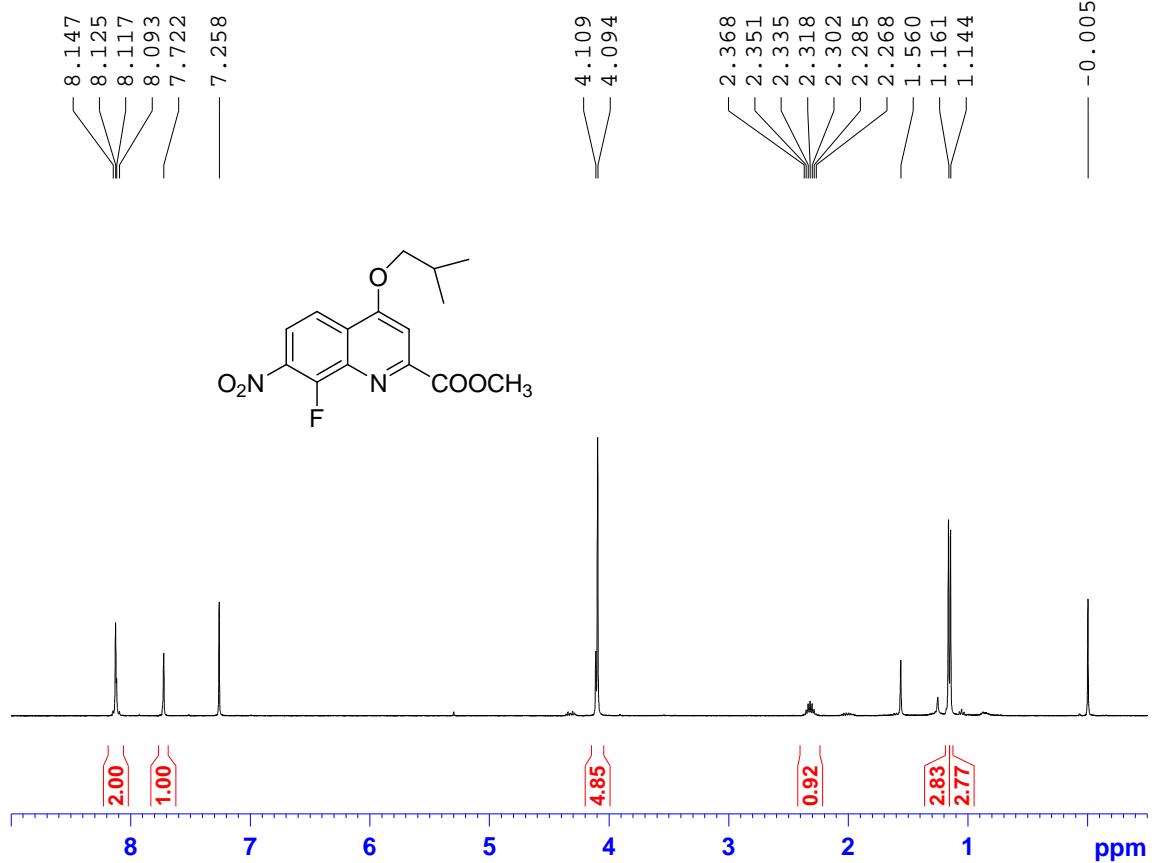
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PULPROG	zg30
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SOLVENT	DMSO
NS	16
DS	0
SWH	11990.407 Hz
FIDRES	0.182959 Hz
AQ	2.7329011 sec
RG	645.1
DW	41.700 usec
DE	6.00 usec
TE	300.5 K
D1	2.0000000 sec
MCREST	0.0000000 sec
MCWRK	0.0150000 sec

===== CHANNEL f1 =====

NUC1	1H
P1	13.50 usec
PL1	-1.00 dB
SFO1	400.1332010 MHz

F2 - Processing parameters

SI	32768
SF	400.1300040 MHz
WDW	EM
SSB	0
LB	0.30 Hz
GB	0
PC	1.00



Current Data Parameters

NAME	gf-22
EXPNO	15
PROCNO	1

F2 - Acquisition Parameters

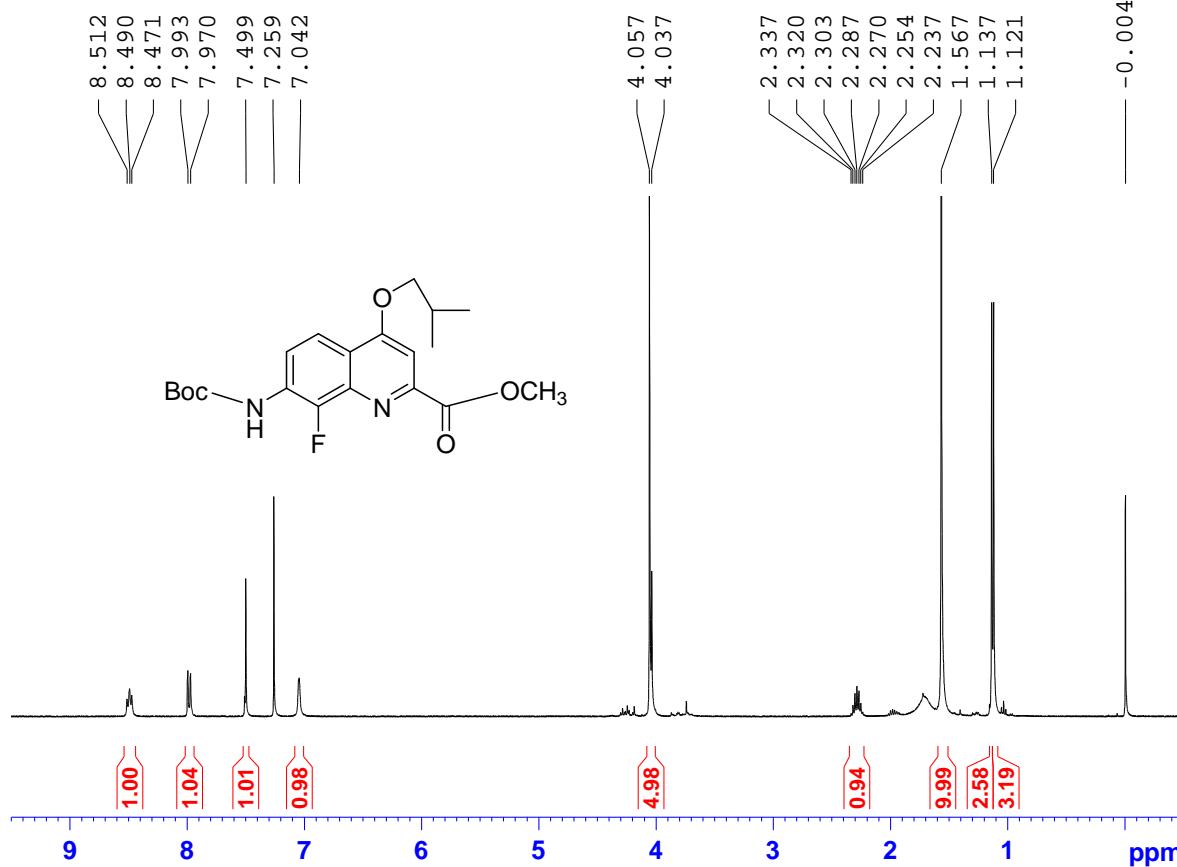
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Time	11.06
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PULPROG	zg30
TD	32768
SOLVENT	CDC13
NS	8
DS	0
SWH	9980.040 Hz
FIDRES	0.304567 Hz
AQ	1.6417269 sec
RG	1149.4
DW	50.100 usec
DE	6.00 usec
TE	294.9 K
D1	2.00000000 sec
MCREST	0.00000000 sec
MCWRK	0.01500000 sec

===== CHANNEL f1 =====

NUC1	1H
P1	8.00 usec
PL1	-4.00 dB
SFO1	400.1326091 MHz

F2 - Processing parameters

SI	32768
SF	400.1300094 MHz
WDW	EM
SSB	0
LB	0.30 Hz
GB	0
PC	1.00

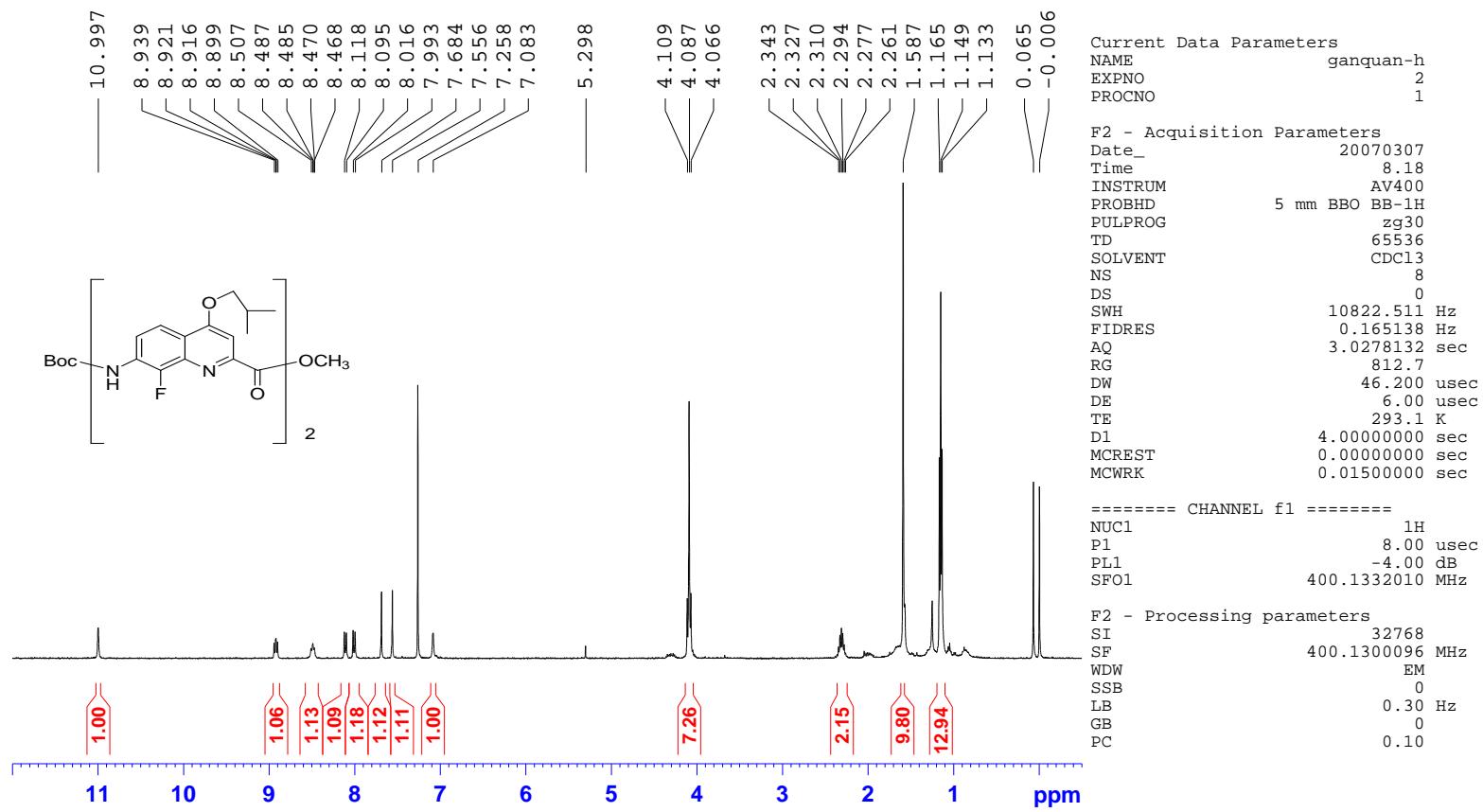


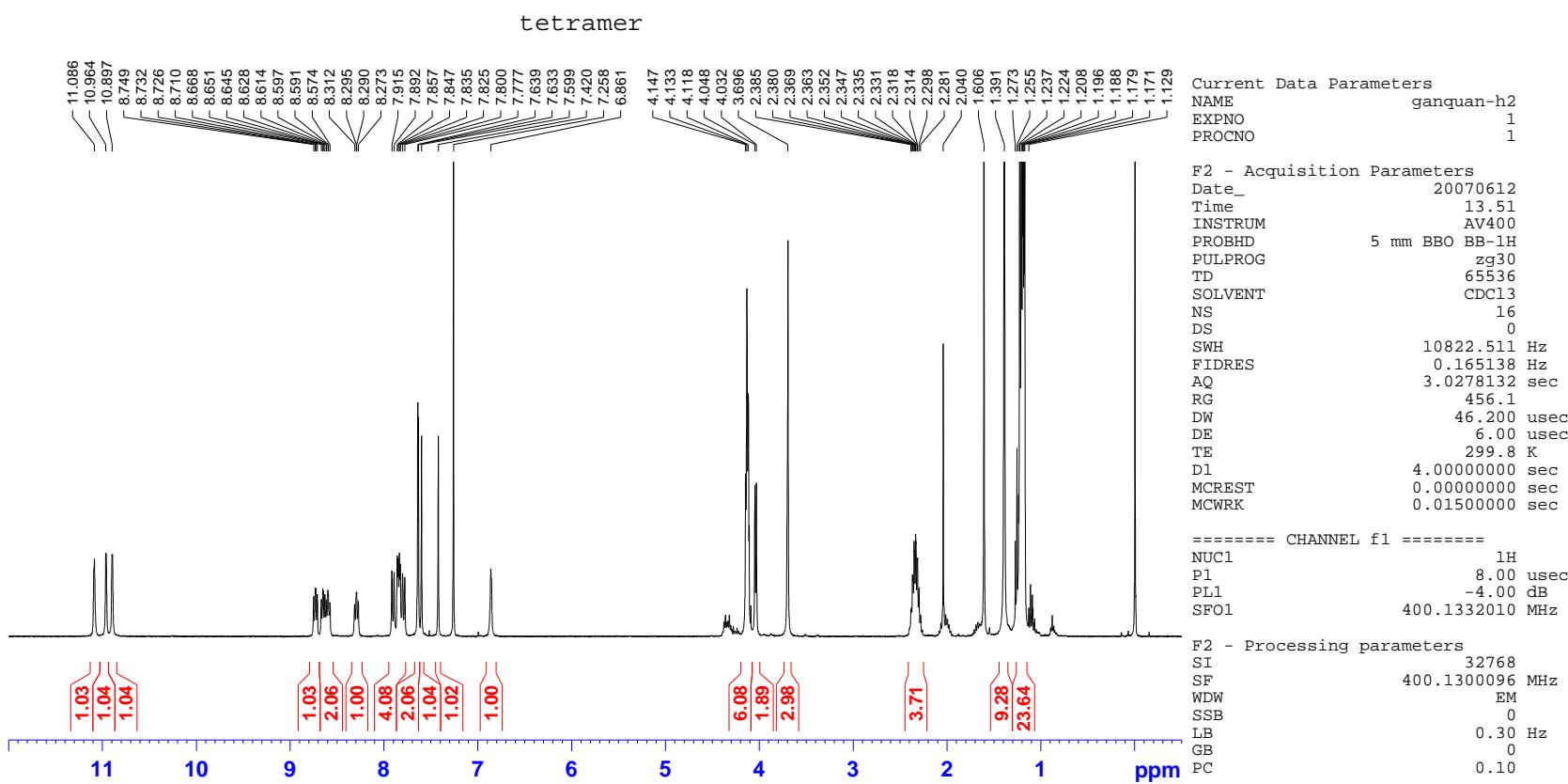
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EXPNO 77  
PROCNO 1

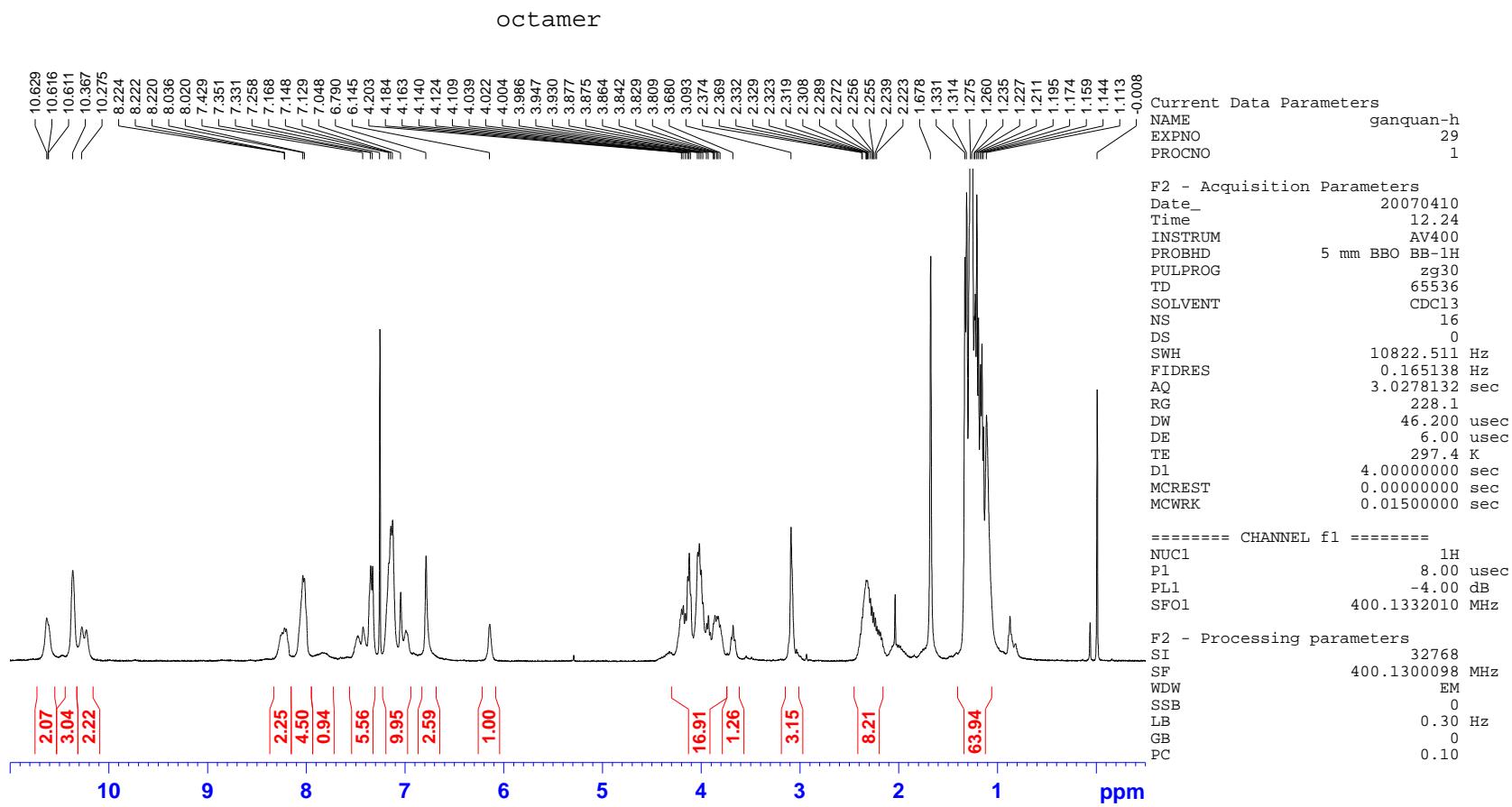
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PROBHD 5 mm PABBO BB-  
PULPROG zg30  
TD 65536  
SOLVENT CDCl3  
NS 16  
DS 0  
SWH 11990.407 Hz  
FIDRES 0.182959 Hz  
AQ 2.7329011 sec  
RG 574.7  
DW 41.700 usec  
DE 6.00 usec  
TE 300.2 K  
D1 2.00000000 sec  
MCREST 0.00000000 sec  
MCWRK 0.01500000 sec

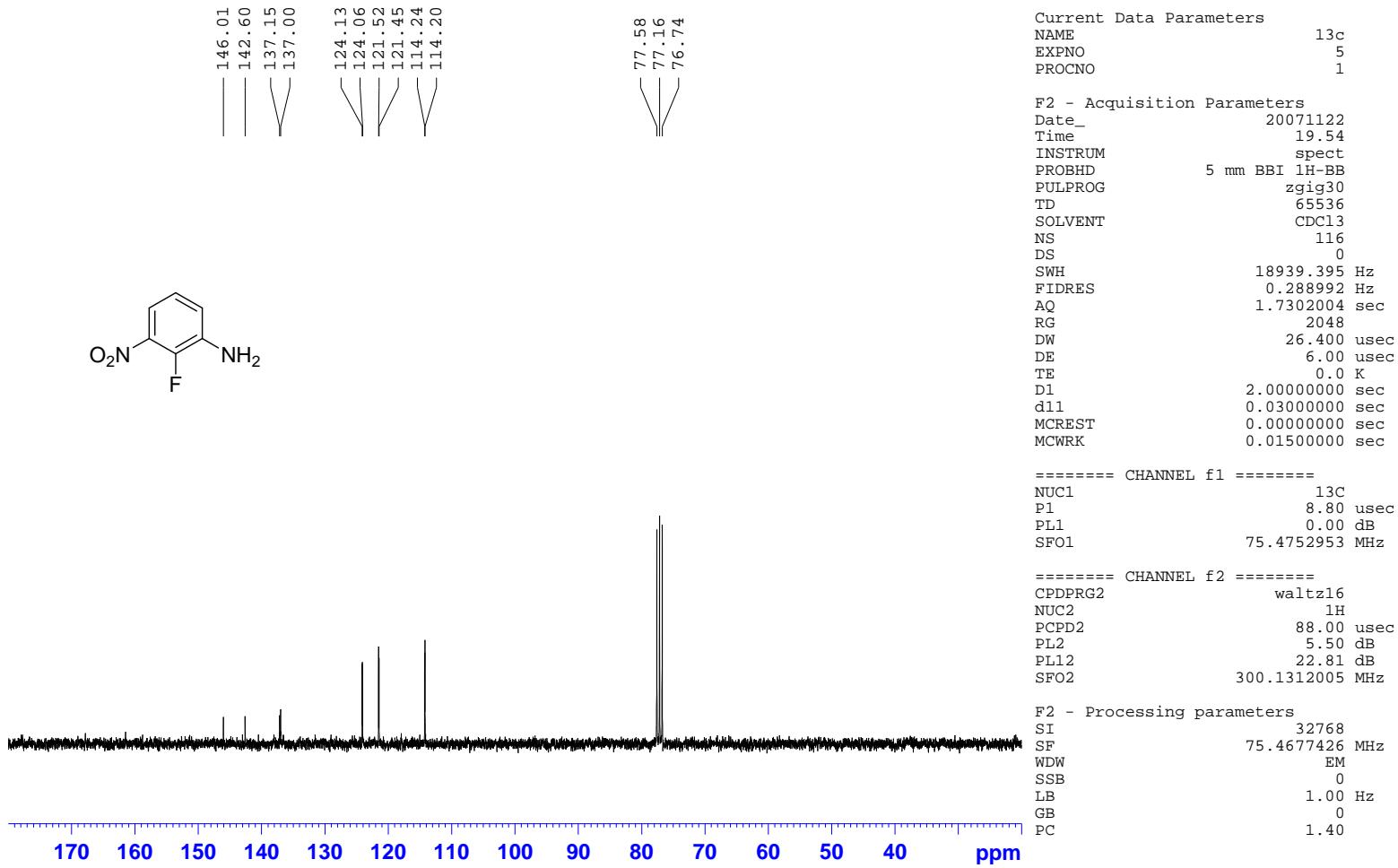
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PL1 -1.00 dB  
SFO1 400.1332010 MHz

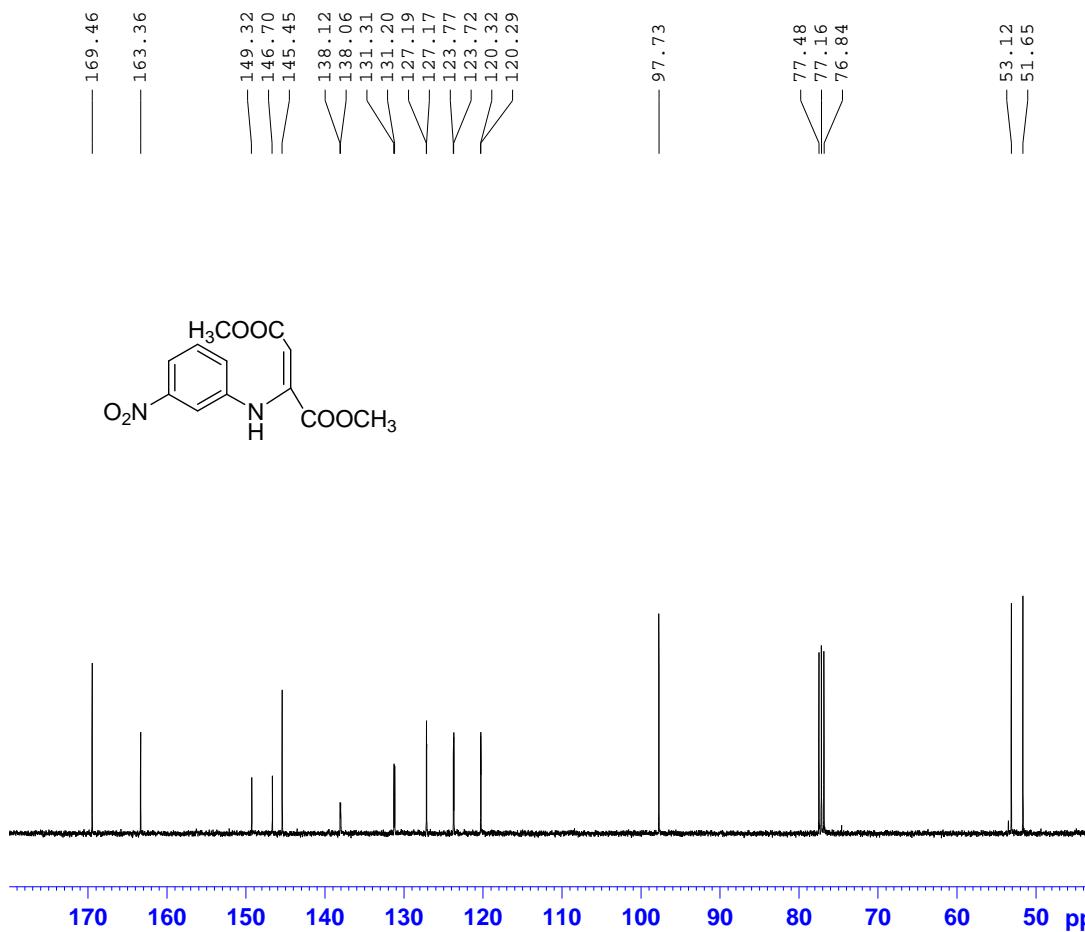
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WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00











Current Data Parameters

NAME	13c
EXPNO	18
PROCNO	1

F2 - Acquisition Parameters

Date_	20071123
Time	12.31
INSTRUM	AV400
PROBHD	5 mm PABBO BB-
PULPROG	zgig30
TD	65536
SOLVENT	CDCl3
NS	90
DS	0
SWH	24154.590 Hz
FIDRES	0.368570 Hz
AQ	1.3566452 sec
RG	14596.5
DW	20.700 usec
DE	6.00 usec
TE	295.7 K
D1	2.0000000 sec
d11	0.0300000 sec
MCREST	0.0000000 sec
MCWRK	0.0150000 sec

===== CHANNEL f1 =====

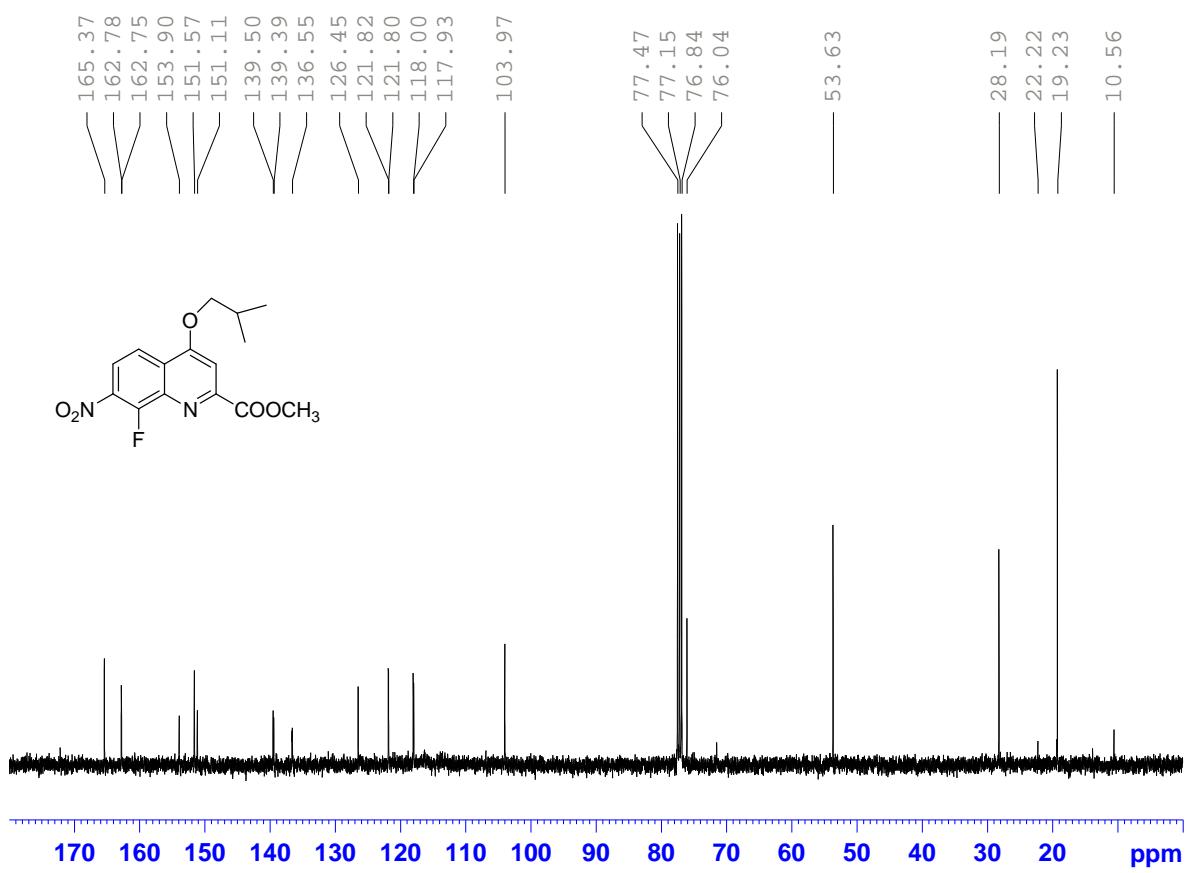
NUC1	13C
P1	9.60 usec
PL1	-3.00 dB
SFO1	100.6238364 MHz

===== CHANNEL f2 =====

CPDPRG2	waltz16
NUC2	1H
PCPD2	80.00 usec
PL2	-1.00 dB
PL12	14.46 dB
SFO2	400.1316005 MHz

F2 - Processing parameters

SI	32768
SF	100.6127667 MHz
WDW	EM
SSB	0
LB	1.00 Hz
GB	0
PC	1.40



Current Data Parameters

NAME	13c
EXPNO	3
PROCNO	1

F2 - Acquisition Parameters

Date	20070706
Time	13.14
INSTRUM	AV400
PROBHD	5 mm BBI 1H-BB
PULPROG	zgig30
TD	65536
SOLVENT	CDCl3
NS	333
DS	0
SWH	23980.814 Hz
FIDRES	0.365918 Hz
AQ	1.3664756 sec
RG	14596.5
DW	20.850 usec
DE	6.00 usec
TE	300.6 K
D1	2.00000000 sec
d11	0.03000000 sec
MCREST	0.00000000 sec
MCWRK	0.01500000 sec

===== CHANNEL f1 =====

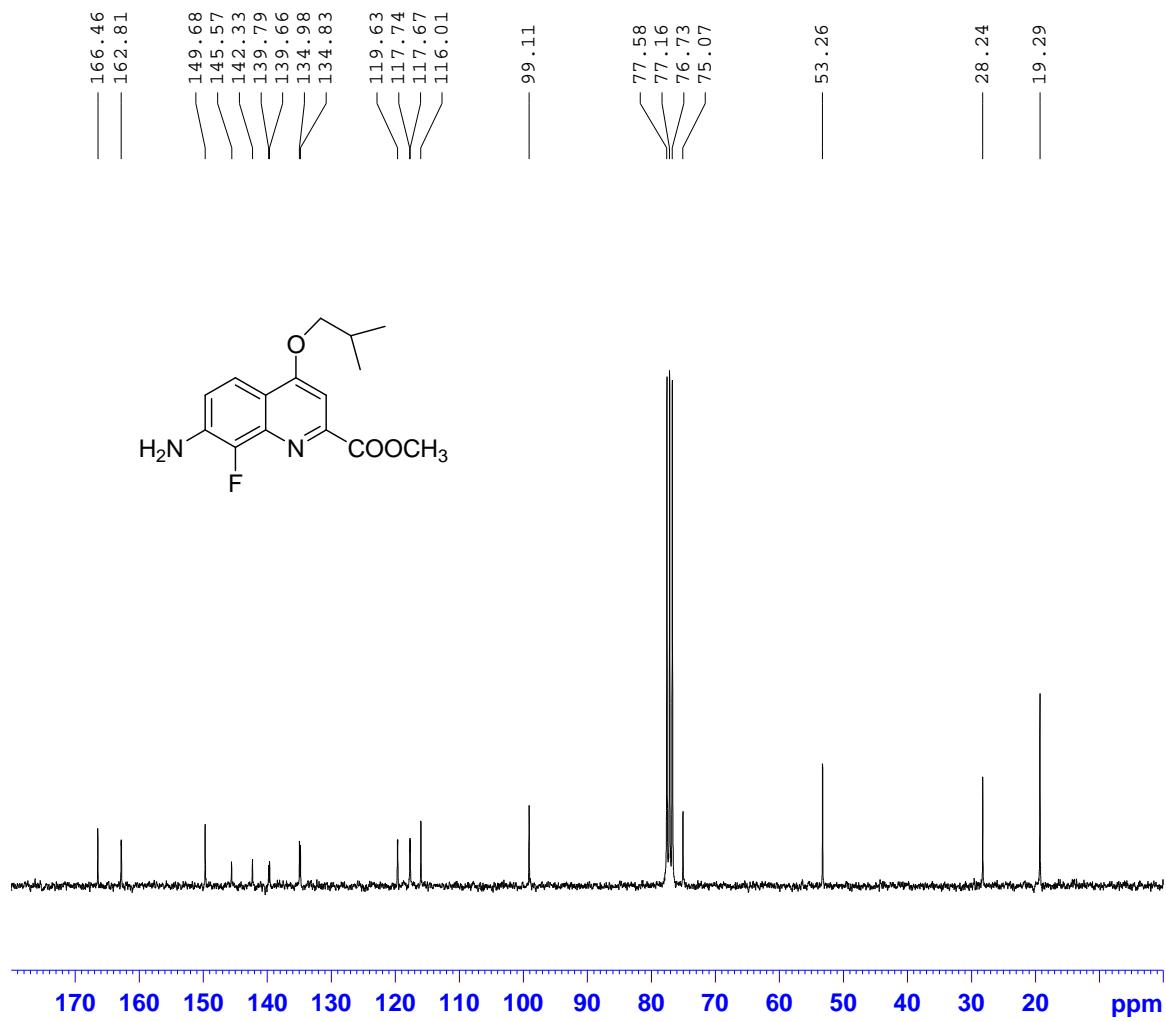
NUC1	13C
P1	21.30 usec
PL1	-3.00 dB
SFO1	100.6228298 MHz

===== CHANNEL f2 =====

CPDPRG2	waltz16
NUC2	1H
PCPD2	80.00 usec
PL2	2.00 dB
PL12	21.27 dB
SFO2	400.1316005 MHz

F2 - Processing parameters

SI	32768
SF	100.6127583 MHz
WDW	EM
SSB	0
LB	1.00 Hz
GB	0
PC	1.40



### Current Data Parameters

NAME	13c
EXPNO	1
PROCNO	1

```

F2 - Acquisition Parameters
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Time            8.25
INSTRUM         spect
PROBHD          5 mm BBI 1H-BB
PULPROG        zgig30
TD              65536
SOLVENT         CDC13
NS              937
DS              0
SWH             18939.395 Hz
FIDRES         0.288992 Hz
AQ              1.7302004 sec
RG              2048
DW              26.400 usec
DE              6.00 usec
TE              0.0 K
D1              2.0000000 sec
d11             0.03000000 sec
MCREST          0.00000000 sec
MCWRK           0.01500000 sec

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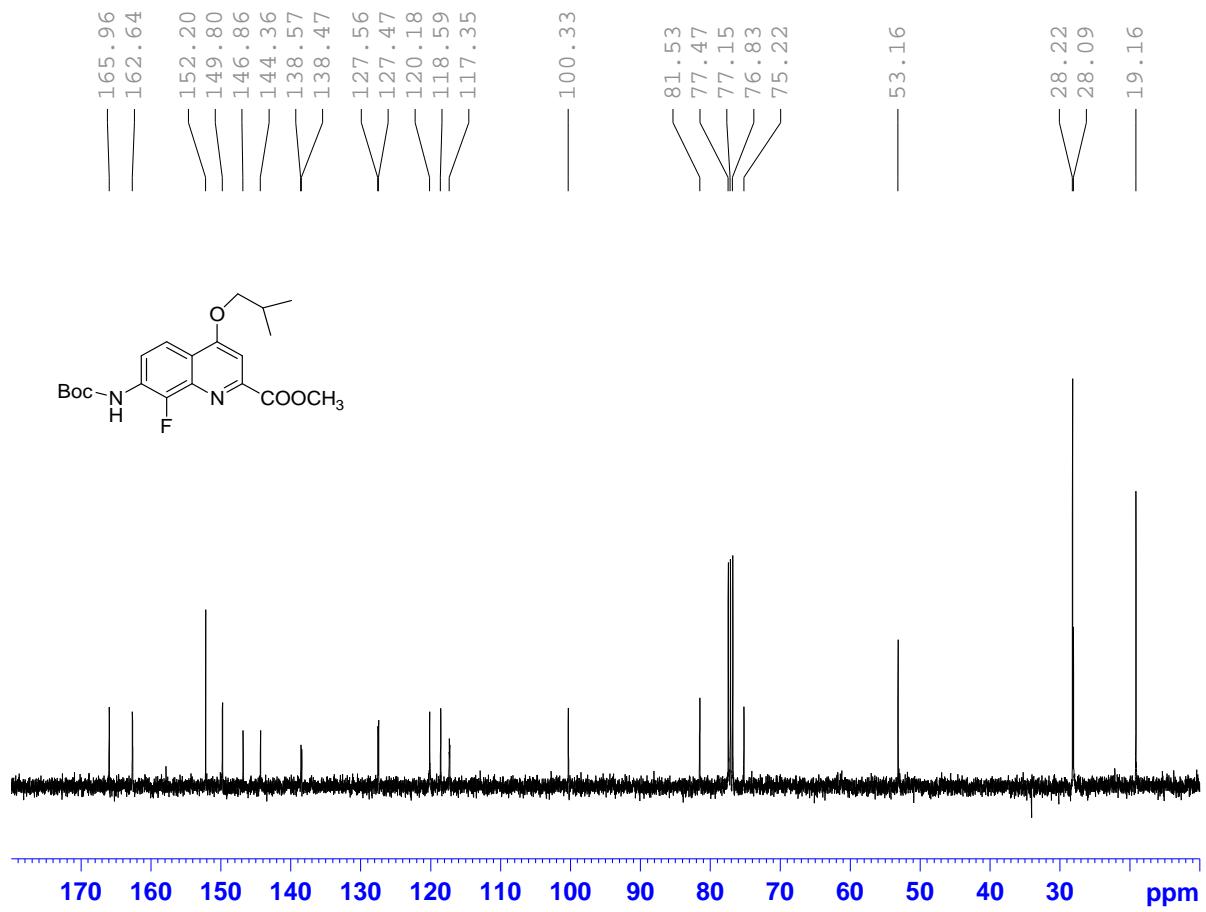
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P1            8.80  usec  
PL1           0.00  dB  
SFO1        75.4752953 MHz
```

```
===== CHANNEL f2 =====
CPDPRG2                      waltz16
NUC2                           1H
PCPD2                      88.00 usec
PL2                           5.50 dB
PL12                         22.81 dB
SF02                        300.1312005 MHz
```

```

F2 - Processing parameters
SI                      32768
SF                      75.4677431 MHZ
WDW                     EM
SSB                     0
LB                      3.00 Hz
GB                     0
PC                     1.40

```



Current Data Parameters  
 NAME 13c  
 EXPNO 13  
 PROCNO 1

F2 - Acquisition Parameters  
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 Time 19.42  
 INSTRUM AV400  
 PROBHD 5 mm PABBO BB-  
 PULPROG zgig30  
 TD 65536  
 SOLVENT CDCl3  
 NS 480  
 DS 0  
 SWH 24154.590 Hz  
 FIDRES 0.368570 Hz  
 AQ 1.3566452 sec  
 RG 14596.5  
 DW 20.700 usec  
 DE 6.00 usec  
 TE 301.9 K  
 D1 2.00000000 sec  
 d11 0.03000000 sec  
 MCREST 0.00000000 sec  
 MCWRK 0.01500000 sec

===== CHANNEL f1 =====  
 NUC1 13C  
 P1 9.60 usec  
 PL1 -3.00 dB  
 SFO1 100.6238364 MHz

===== CHANNEL f2 =====  
 CPDPRG2 waltz16  
 NUC2 1H  
 PCPD2 80.00 usec  
 PL2 -1.00 dB  
 PL12 14.46 dB  
 SFO2 400.1316005 MHz

F2 - Processing parameters  
 SI 32768  
 SF 100.6127663 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40

